



دانشگاه صنعتی اصفهان

صنایع لبنی تکمیلی

دکتر علی نصیرپور

1391



تعداد واحد: ۲

نوع واحد: نظری

پیشنیاز: ندارد

سرفصل درس:

اثر عملیات حرارتی - اصول اساسی، ستیک و واکنشها طی عملیات حرارتی - اثر حرارت بر میکروارگانیسمها و آنزیمها، اثر حرارت بر نمکها و PH شیر، واکنش میلارد، عوامل موثر بر قهوه‌ای شدن - تشخیص آزمایشگاهی شیرهای استریل در ظروف و روشهای UHT - اثر حرارت بر پروتئینهای شیر - پروتئینهای سرمی - پیامدهای دناتوراسیون پروتئینهای سرمی - واکنشهای گوگرد موجود در پروتئینها - پایداری محلولهای کلوئیدی پروتئین در حین عملیات حرارتی - عوامل موثر بر پایداری حرارتی - تغییرات فیزیکی و شیمیایی تغییر طعم - تغییر خاصیت اکسیداتیو - تغییر رنگ - رویه بستن شیر - تغییر ارزش تغذیه‌ای - شیر به عنوان محیط کشت برای پرورش استارترها - خاصیت کلوئیدی و پدیده سطحی - شیمی سطحی - پایداری کلوئیدی - تغییرات انتشار - پراکندگی (اندازه) میل کازئین - خصوصیات پایداری کلوئیدی - اثر آنزیم رنتین - بسته شدن پیری - مقاومت حرارتی گریچه‌های چربی - خصوصیات پایداری امولسیون - عکس العمل در مقابل سرما - (اگلوتیناسیون سرما) - هموژنیزاسیون - خامه‌ای شدن - اثر متقابل ترکیبات شیر با هوا - تئوریهای کف کردن - کف کردن فرآورده‌های لبنی - چرینگ (زدن)، خصوصیات رئولوژیکی - رفتار نیوتنی - محلولهای غیر نیوتنی - ژل و چربی شیر. خصوصیات ترکیبات تغلیظ شده شیر - عکس العمل آب - اثر تغلیظ / تبخیر - انجماد - خشک کردن - فرایند ممبران - تغلیظ پروتئین - بازسازی / طعم و آروما - خصوصیات کلی - آرومای شیر / آرومای مختلف لبنی - تغییرات آروما در شیر و فرآورده‌های لبنی .

References:

- 1- Dairy processing handbook. 1995-2005. Tetrapak.
- 2- Dairy science and technology. 2006. Walstra et al
- 3- Dairy Science and Technology Handbook. 1993. YH HUI
- 4- Dairy Chemistry. 1997. Fox
- 5- Dairy Chemistry and Biochemistry. 1998. Fox and McSWEENEY.
- 6- Dairy processing Improving quality. 2000. Gerrit Smit. CRC Press.

هدف از فرایند حرارتی:

1- تضمین سلامت مصرف کننده:

Coxiella burnetii

Mycobacterium tuberculosis

Staphylococcus aureus

Listeria monocytogenes,

Salmonella species

Campylobacter jejuni

پاتوژن های مقاوم به حرارت ممکن است:

- در شیر وجود نداشته باشند (*Bacillus anthracis*)،
- یا به دلیل رشد سایر گونه ها فرصت رشد نمی یابند (*Clostridium perfringens*)،
- یا در شیر رشد نمی کنند (*Clostridium botulinum*)،
- یا در تعداد زیاد ایجاد مسمومیت یا بیماری می نمایند (*Bacillus cereus*)

2- افزایش عمر نگهداری

3- ایجاد خصوصیات ویژه در برخی محصولات

CHANGES CAUSED BY HEATING:

➤ **Reversible or irreversible**

Reversible: mutarotation equilibrium of lactose and changes in ionic equilibriums, including pH

Chemical and Physical Changes:

- Gases, including CO₂, are partly removed
- The amount of colloidal phosphate increases and the [Ca²⁺] decreases
- Lactose isomerizes and partly degrades to yield, for instance, lactulose and organic acids
- milk pH decreases
- Most of the serum proteins are denatured
- Part of the serum protein (especially of β-lactoglobulin) becomes covalently bound to κ-casein and to some proteins of the fat globule membrane.
- Enzymes are inactivated
- Reactions between protein and lactose occur, Maillard reactions in particular
- Casein micelles become aggregated. Aggregation may eventually lead to coagulation

- Several changes occur in the fat globule membrane, e.g., in its Cu content
- Some vitamins are degraded

Consequences:

- Bacterial growth rate of the organisms surviving, or added after heat treatment, can be greatly affected, generally increased:

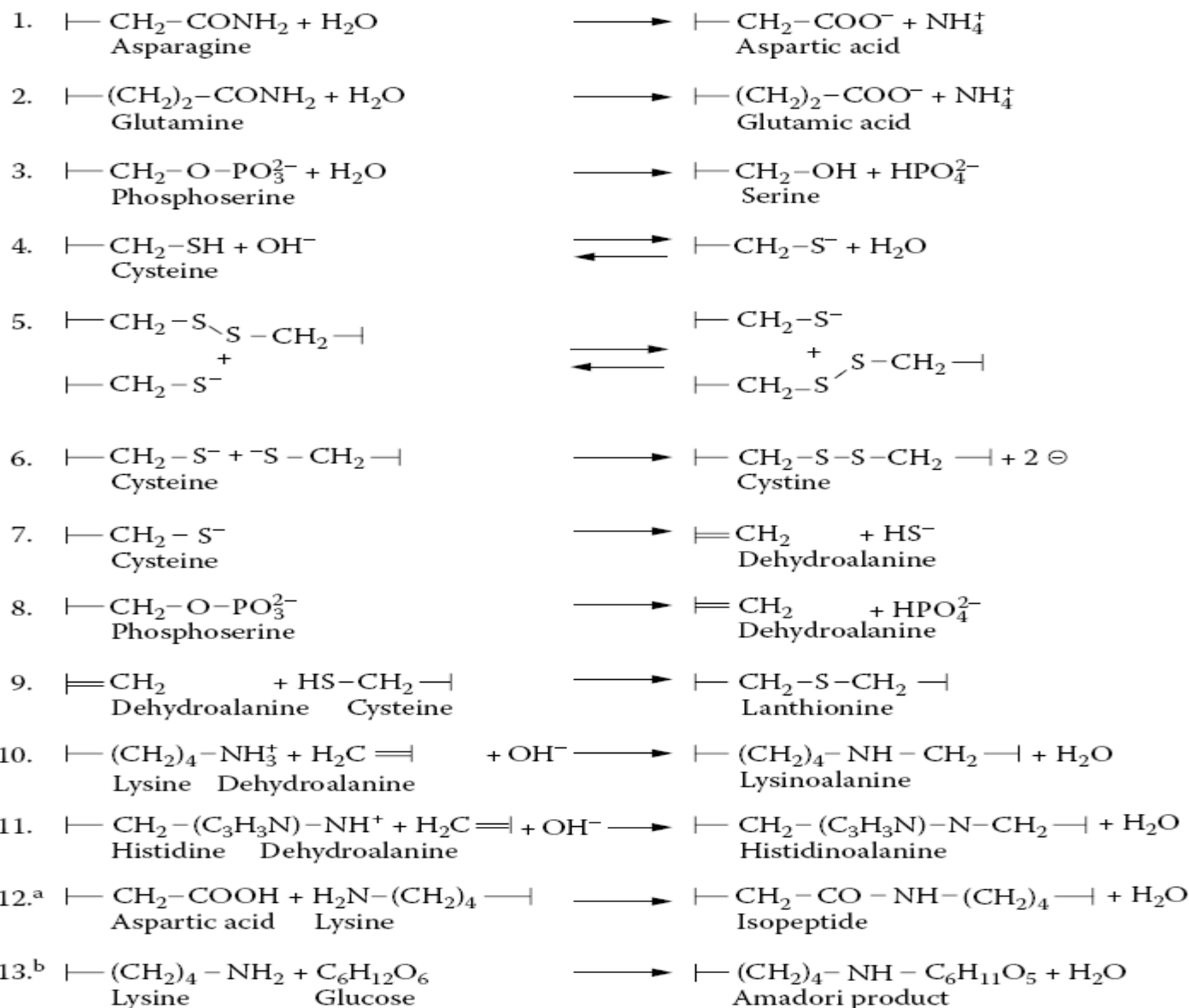
IgM (agglutinin) Bacillus cereus
lactoperoxidase system → lactic acid bacteria
Lactoferrin Bacillus ~~stearo~~thermophilus

Bacteriophages ~~can be~~ inactivated, depending on the heating intensity

- Nutritive value decreases
- The flavor changes appreciably
- Color may change
- Viscosity may increase slightly
- Heat coagulation in evaporated milk serum protein is denatured before concentrating
- Age gelation in sweetened condensed milk ~~is also~~ reduced when the milk is intensely heated before concentrating.

- The *rennetability of milk and the rate of syneresis of the rennet gel* decrease (serum proteins bound to k-casein)
- *Creaming tendency of the milk decreases*

Possible Reactions of Side Chain Groups of Amino Acid Residues Linked in the Peptide Chain (|) of Proteins at High Temperature



^a Reaction also occurs with glutaminic acid residues.

^b First step in the Maillard reaction with glucose or another reducing sugar. See [Figure 7.4](#).

Denaturation of serum proteins

Unfolding in peptide chains (temperature 80 °C): reaction in or between side groups chains preventing → refolding

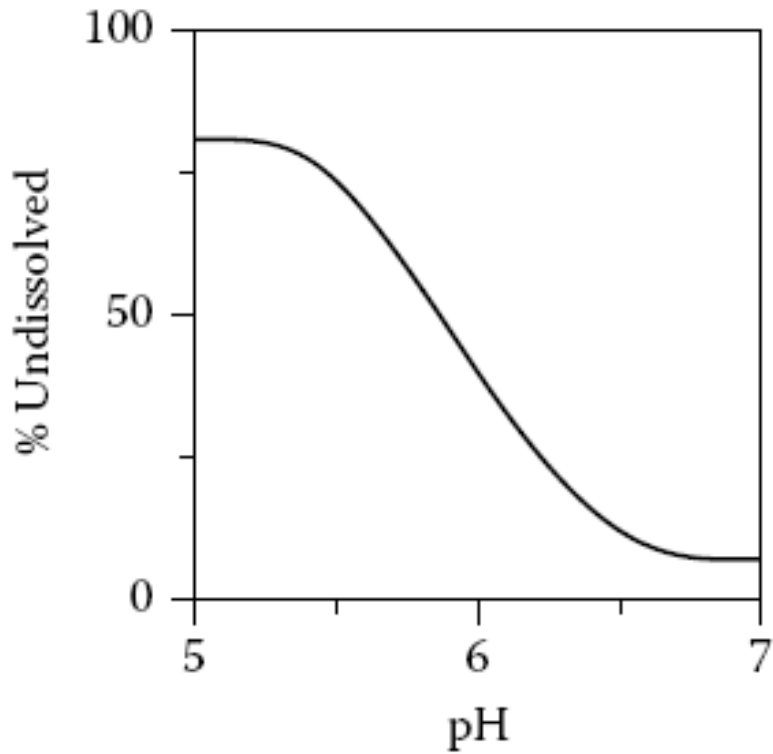
These changes happen for serum proteins specially BLG, ALA, serum albumin, Immunoglobulins.

Proteose peptones do not denatured (caseins)

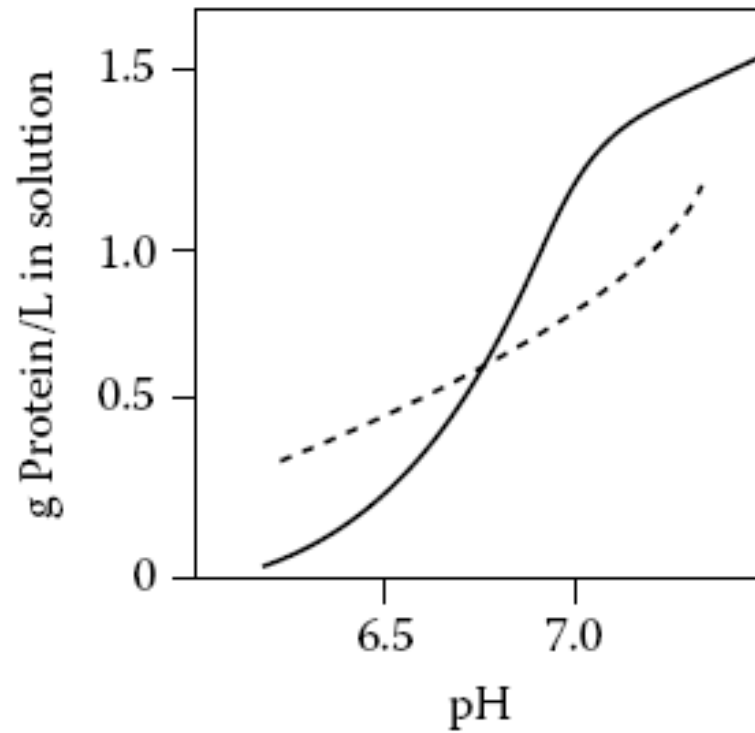
At high temperature, thiol group react with –s-s- groups —Dimer,
trimer, tetramer,

In milk during heating: BLG react with k-casein —casein micells
cover with BLG (depending on pH)

Whey proteins show different thermal stabilities: alpha-lactalbumin > betalactoglobulin > bovine serum albumin > immunoglobulins.



(A)



(B)

Influence of pH on the effects of heating on proteins. (A) Percentage of the proteins that become precipitated after heating whey for 10 min at 80°C. (B) Amount of protein that remains in solution, i.e., not associated with the casein micelles, after heating milk (—) or serum protein free milk (----) at 140°C.

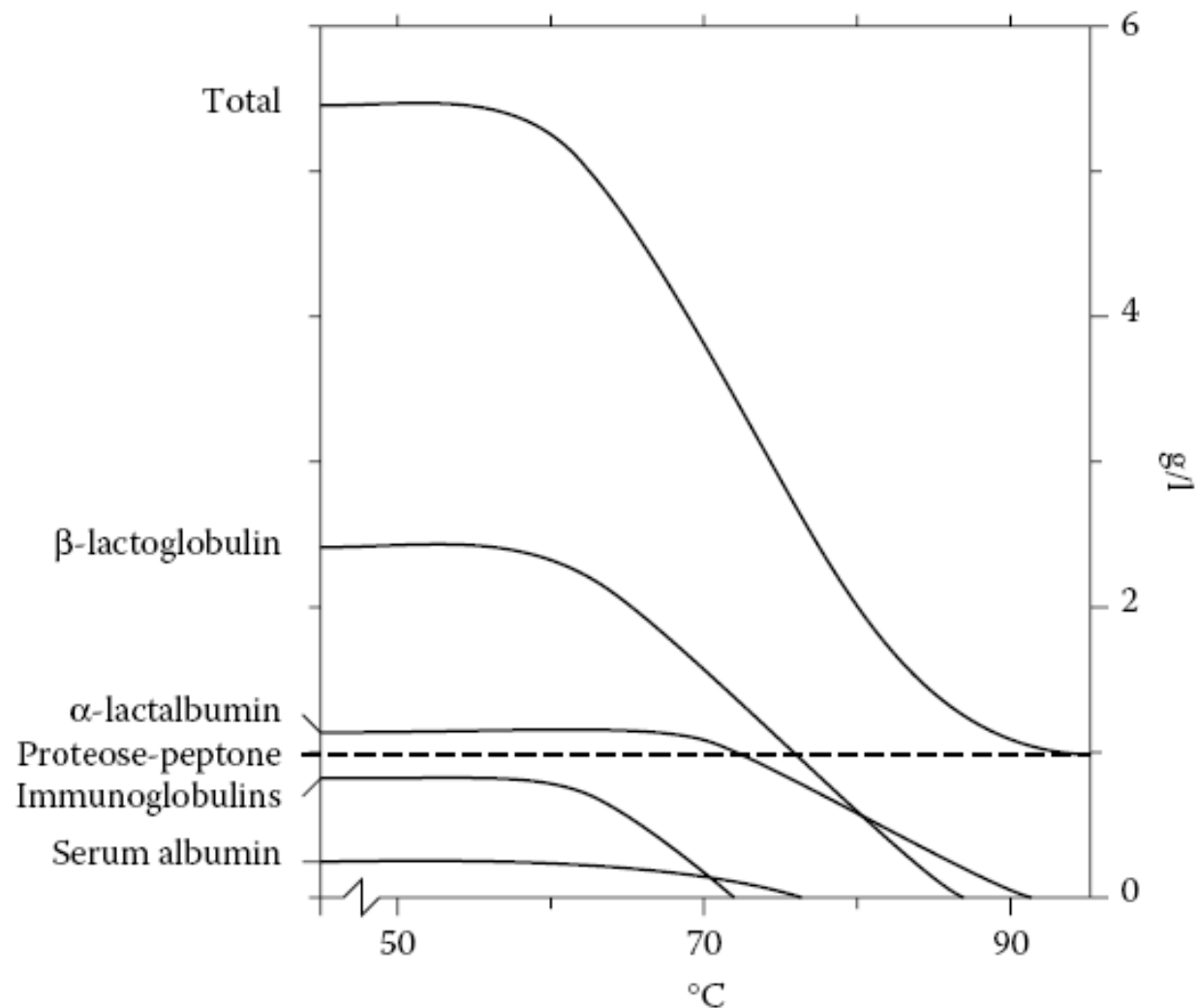


FIGURE 7.3 Effect of heating milk for 30 min at various temperatures on quantity of serum proteins that remain dissolved after cooling and acidification to pH 4.6. (Mainly adapted from B.L. Larson and G.D. Roller, *J. Dairy Sci.*, **38**, 351, 1955.)

ISOMERIZATION AND SUGAR DEGRADATION REACTIONS

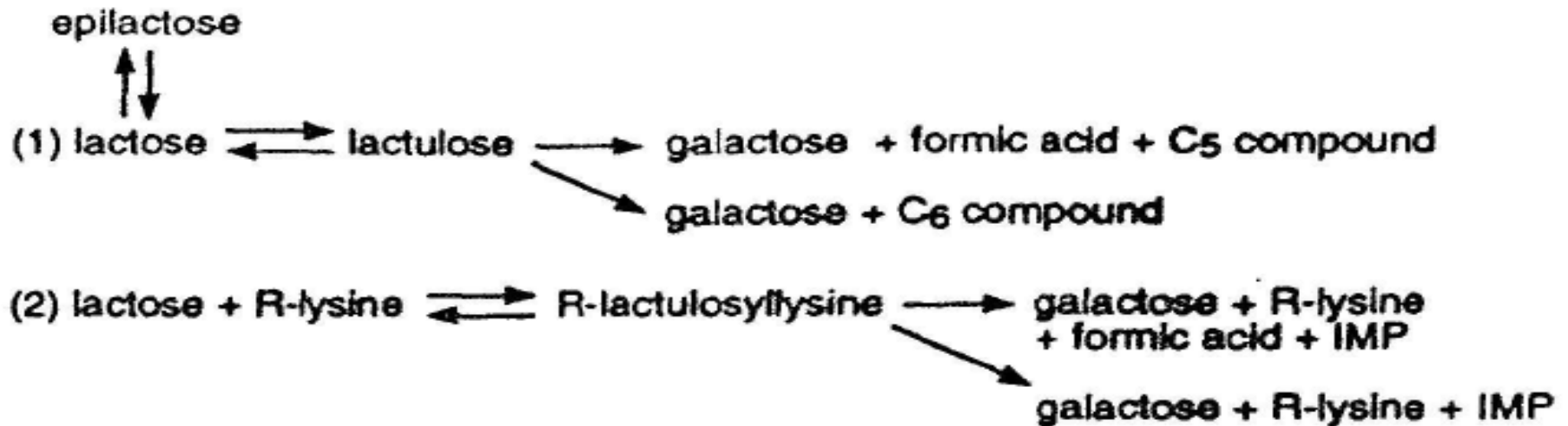
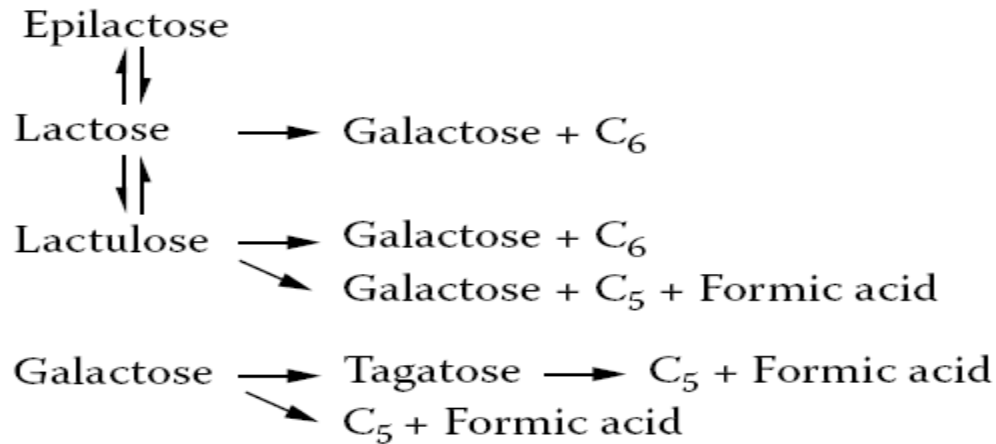
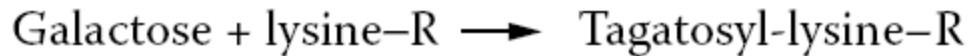
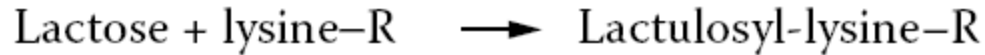


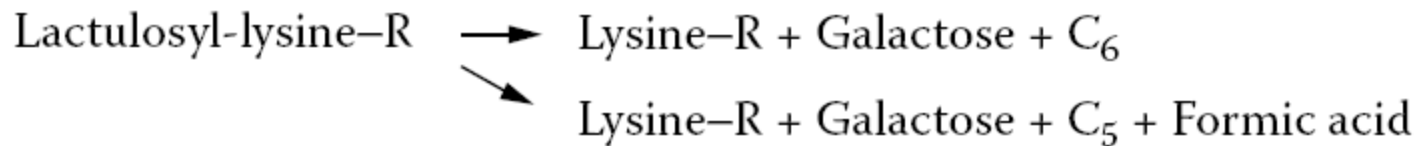
FIGURE 6.5 Simplified scheme of reactions occurring in the initial stage of the breakdown of lactose during the heating of milk (sterilization temperatures). IMP, intermediate Maillard products.

MAILLARD REACTIONS

Initial



Intermediate



Advanced

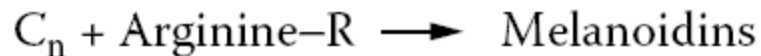
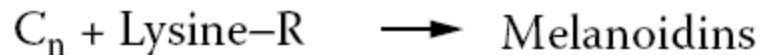


FIGURE 7.4 Simplified scheme of reactions occurring with lactose during the heating of milk at sterilization temperature. R stands for a peptide chain, C_n for an organic compound containing n carbon atoms.

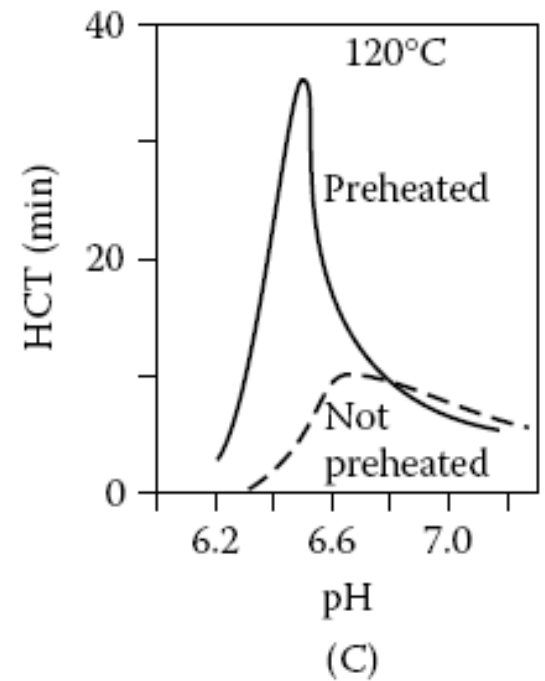
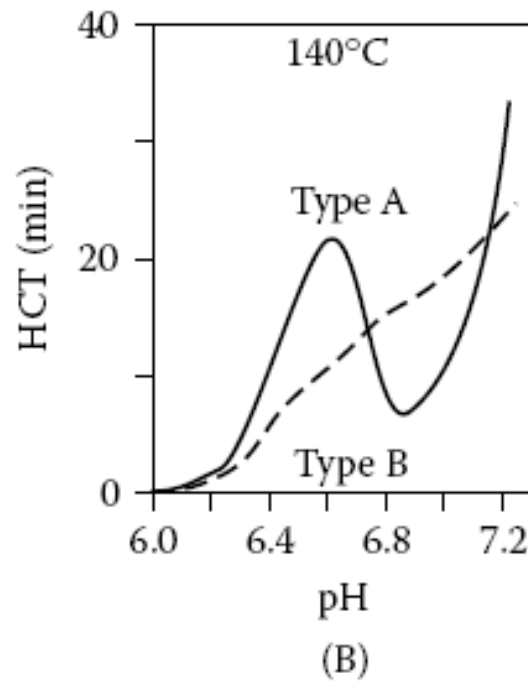
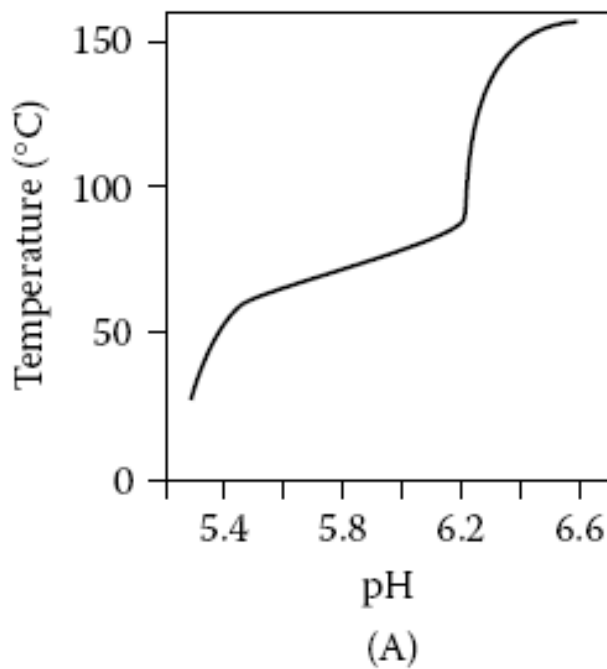


FIGURE 7.6 Heat coagulation of milk as a function of the initial pH. (A) Temperatures at which coagulation starts at fairly rapid warming of the milk (approximate results). (B) Heat coagulation time at 140°C of two different samples of fresh milk. (C) HCT at 120°C of evaporated skim milk, with or without preheating of the milk before concentration.

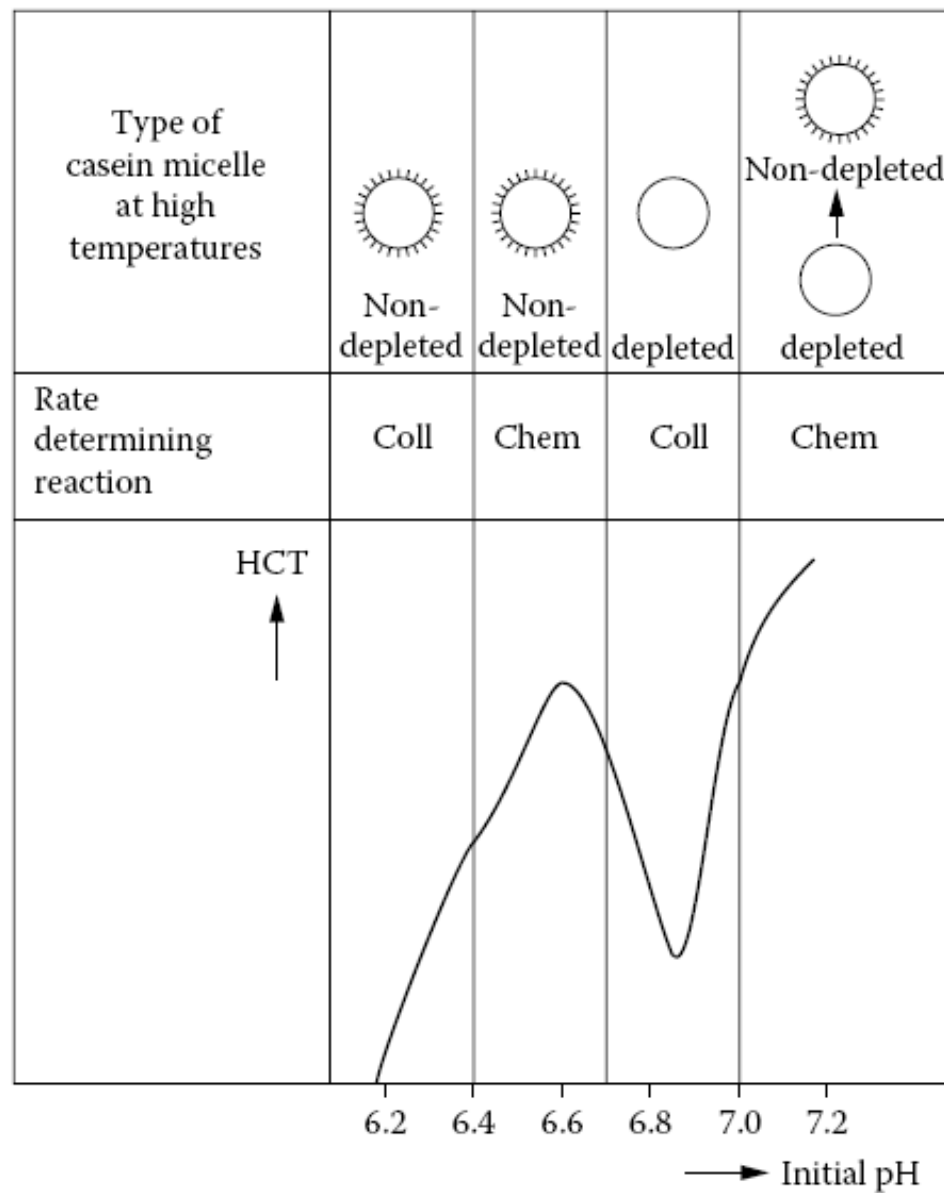


FIGURE 7.7 Model for the effect of initial pH on the type of casein micelle emerging at high temperature and thereby on the heat coagulation time (HCT) of milk. coll = colloidal aggregation, chem = chemical reaction

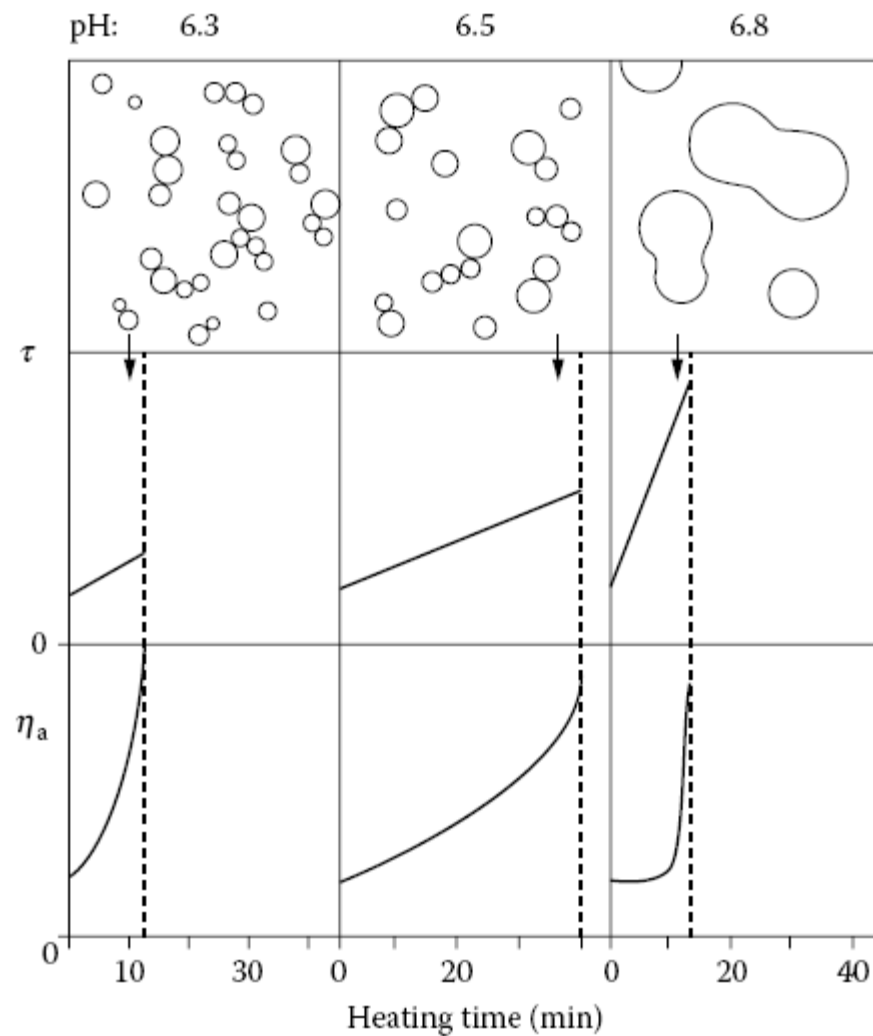


FIGURE 7.8 Heat coagulation at 120°C of concentrated skim milk at various initial pH. The upper row shows the appearance of the casein micelles (derived from electron micrographs) at the moments indicated by arrows, i.e., shortly before heat coagulation. The HCT is indicated by a vertical broken line. The second row gives the turbidity (t) as a function of heating time, the lowest row the apparent viscosity (h_a). t and h_a were determined in situ, i.e., at 120°C. Approximate results.

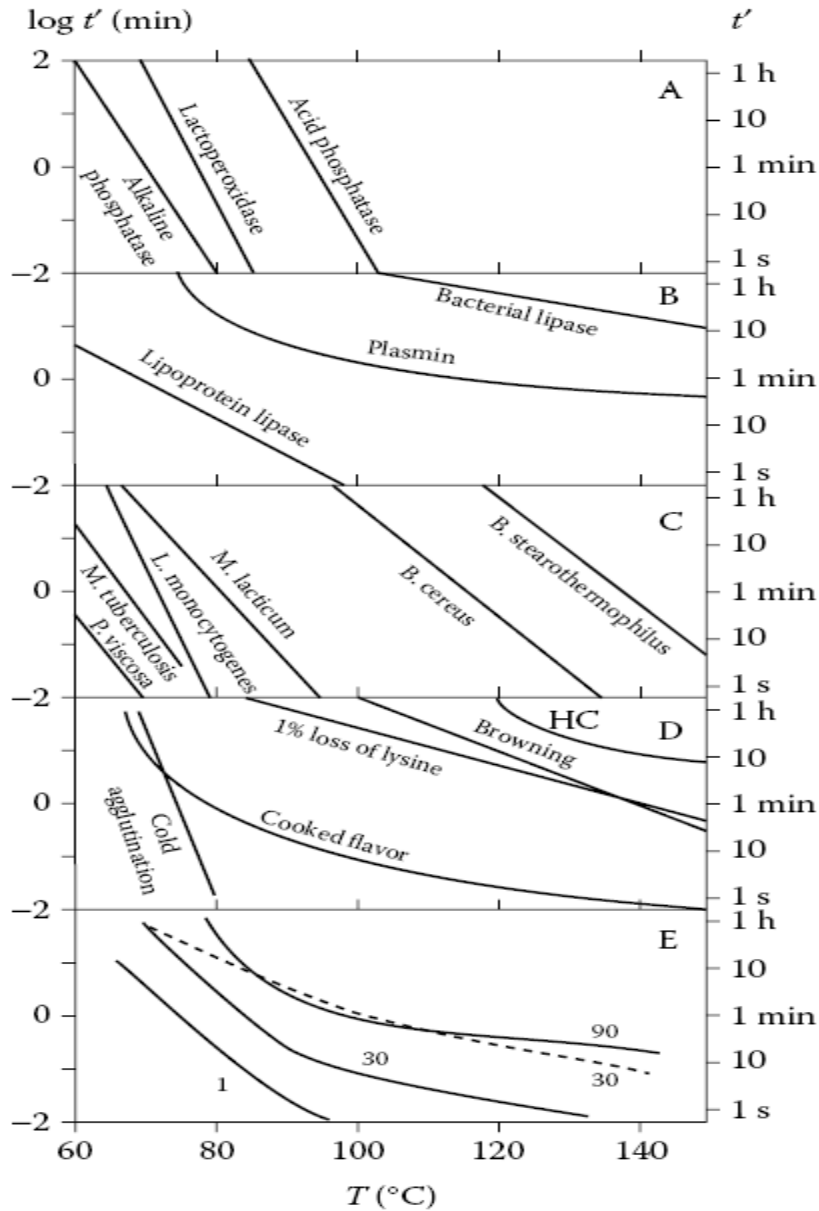


FIGURE 7.9 Combinations of temperature (T) and time (t') of heat treatment of milk that cause (A, B) inactivation (reduction of activity to about 1%) of some milk enzymes and a bacterial lipase; (C) the killing (reduction of the count to 10^{-6}) of strains of the bacteria *Pseudomonas viscosa*, *Mycobacterium tuberculosis*, *Listeria monocytogenes*, and *Microbacterium lacticum*, and of spores (10^{-4}) of *Bacillus cereus* and *B. stearothermophilus*; (D) visible heat coagulation (HC), a certain degree of browning, decrease in available lysine by 1%, a distinct cooked flavor and inactivation of cold agglutination; (E) insolubilization of 1%, 30%, and 90% of the β -lactoglobulin, and of 30% of the α -lactalbumin (----). Approximate results.

Kinetic aspects of milk heat treatment

$$-dc/dt = Kc$$

$$\ln (c_0/c) = Kt$$

$$c = c_0 e^{-Kt}$$

$$t' = \ln (c_0/c')/K$$

$$D = (\ln 10)/K \approx 2.3/K$$

$$dc/dt = K$$

$$c = Kt + c_0$$

$$K(T) = K_0 \exp(-E_a/RT)$$

$$Q_{10} \equiv K(T + 10)/K(T)$$

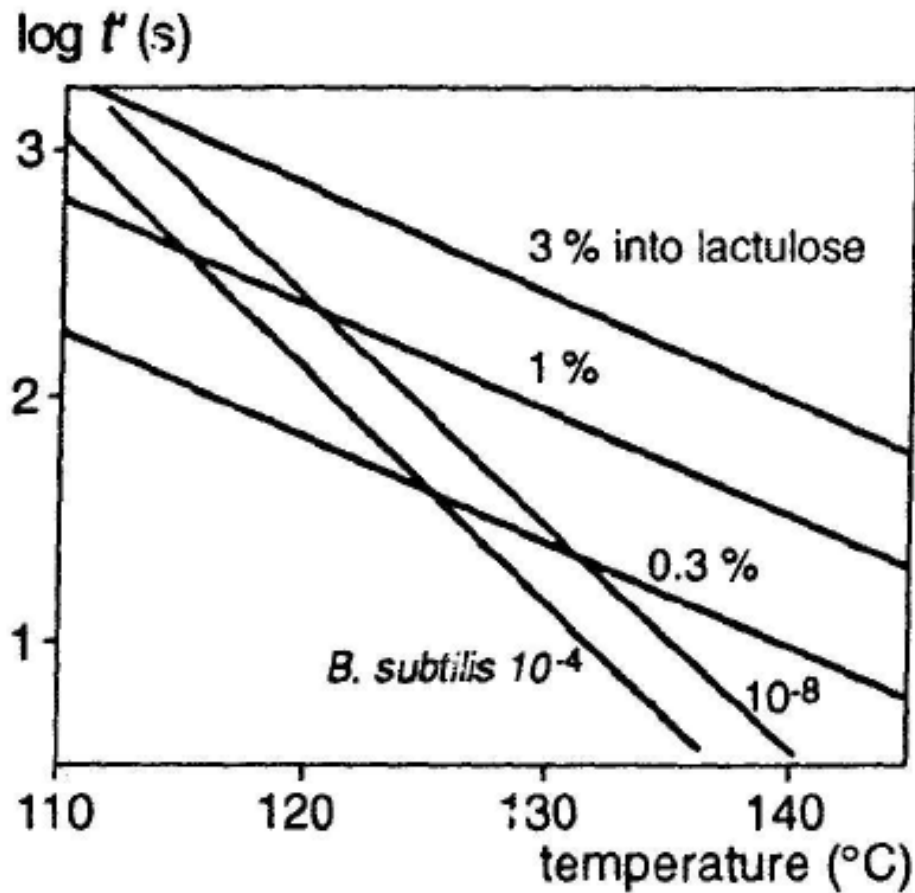


FIGURE 6.10 The time needed (t') at various temperatures to convert certain percentages of lactose to lactulose, and to obtain a certain extent of killing of *Bacillus subtilis* spores.

TABLE 6.3 Typical Examples of the Temperature Dependence of Some Reactions

| Type of reaction | Activation energy ^a (kJ · mol ⁻¹) | Q ₁₀ at 100°C |
|---------------------------------|---|--------------------------|
| Many chemical reactions | 80–125 | 2–3 |
| Many enzyme-catalyzed reactions | 40–60 | 1.4–1.7 |
| Autoxidation of lipids | 40–100 | 1.4–2.4 |
| Maillard reactions | 100–180 | 2.4–5 |
| Heat denaturation of proteins | 200–600 | 6–175 |
| Enzyme inactivation, e.g., | 450 | 50 |
| Killing of vegetative bacteria | 200–600 | 6–175 |
| Killing of spores | 250–330 | 9–17 |

^a Often an apparent or average activation energy because it mostly concerns a number of different ensuing reactions.

TABLE 6.4 Killing of Some Bacteria Due to Heating

| | Heating medium | Temp. (°C) | D (min) | Z (K) |
|---|----------------|------------|-----------|---------|
| Psychrotrophs | | | | |
| <i>Pseudomonas fragi</i> | Milk | 49 | 7-9 | 10-12 |
| <i>Pseudomonas fragi</i> | Skim milk | 49 | 8-10 | 10-12 |
| <i>Pseudomonas fragi</i> | Whey, pH 6.6 | 49 | 32 | |
| <i>Pseudomonas fragi</i> | Whey, pH 4.6 | 49 | 4-6 | 10.9 |
| <i>Pseudomonas viscosa</i> | Milk | 49 | 1.5-2.5 | 4.9-7.9 |
| <i>Pseudomonas viscosa</i> | Whey, pH 6.6 | 49 | 3.9 | |
| <i>Pseudomonas viscosa</i> | Whey, pH 4.6 | 49 | 0.5 | |
| <i>Pseudomonas fluorescens</i> | Buffer | 60 | 3.2 | 7.5 |
| <i>Microbacterium thermosphaerum</i> | Skim milk | 50 | 2.5 | |
| <i>Listeria monocytogenes</i> | Milk | 65 | 0.1 | 6.6 |
| <i>Listeria monocytogenes</i> | Skim milk | 72 | 0.07 | 6.5 |
| <i>Yersinia enterocolitica</i> | Milk | 62.8 | 0.01-0.3 | |
| Other non-spore-forming bacteria | | | | |
| <i>Salmonella</i> (6 spp.) | Milk | 62.8 | 1.5-4.5 | 4.0-5.2 |
| <i>Salmonella</i> (2 spp.) | Milk chocolate | 62.8 | 1100-1950 | 18-19 |
| <i>Staphylococcus aureus</i> | Milk | 62.8 | 7-30 | 5.0-5.2 |
| <i>Campylobacter jejuni</i> | Milk | 50 | 3.5-5.5 | 6-8 |
| <i>Escherichia coli</i> | Skim milk | 62.8 | 0.13 | 4.6 |
| <i>Escherichia coli</i> | Whey, pH 4.6 | 62.8 | 0.26 | 6.7 |
| <i>Streptococcus</i> sp., group D | Skim milk | 62.8 | 2.6 | |
| <i>Streptococcus faecalis</i> | Skim milk | 62.8 | 3.5 | |
| <i>Streptococcus faecium</i> | Skim milk | 62.8 | 10.3 | |
| <i>Streptococcus durans</i> | Skim milk | 62.8 | 7.5 | |
| <i>Streptococcus bovis</i> | Skim milk | 62.8 | 2.6 | |

| | | | | |
|--|--------------------------|------|-------------|---------|
| <i>Lactococcus lactis</i> ssp. <i>lactis</i> | Whey, pH 4.6 | 62.8 | 0.32 | 7.3 |
| <i>Lactococcus lactis</i> ssp. <i>cremoris</i> | Whey, pH 4.6 | 62.8 | 0.036 | 6.7 |
| <i>Lactobacillus</i> spp. | Milk | 65 | 0.5-2.0 | |
| <i>Microbacterium flavum</i> | Skim milk | 65 | 2.0 | |
| <i>Microbacterium lacticum</i> | Skim milk | 70 | 4.0 | |
| Spore-forming bacteria | | | | |
| <i>Bacillus cereus</i> , spores | Milk | 121 | 0.04 | 9.4-9.7 |
| <i>Bacillus cereus</i> , vegetative | Water or 2 M sucrose | 70 | 0.013-0.016 | |
| <i>Bacillus cereus</i> , germinating spore | Water | 70 | 0.35 | |
| <i>Bacillus cereus</i> , germinating spore | 2 M sucrose | 70 | 39 | |
| <i>Bacillus licheniformis</i> | Skim milk, pH 6.7 | 111 | 0.48 | 8 |
| <i>Bacillus licheniformis</i> | Skim milk, pH 6.3 | 111 | 0.35 | 8 |
| <i>Bacillus subtilis</i> , spore | Milk | 121 | 0.03-0.5 | 10.7 |
| <i>Bacillus subtilis</i> , vegetative | Water | 55 | 1.0-5.6 | |
| <i>Bacillus subtilis</i> , vegetative | 2 M sucrose | 55 | 0.8-62 | |
| <i>Bacillus coagulans</i> , spore | Milk | 121 | 0.6-4 | |
| <i>Bacillus pumilus</i> , spore | Milk | 95 | 1.4 | 9.7 |
| <i>Bacillus stearothermophilus</i> , spore | Milk | 121 | 4-7 | 8-11 |
| <i>Clostridium sporogenes</i> , spore | Milk, pH 7.0 | 121 | 1.7 | |
| <i>Clostridium botulinum</i> , spore | Milk, pH 7.0 | 121 | 0.2 | |
| <i>Clostridium botulinum</i> , type A, spore | Phosphate buffer, pH 7.0 | 110 | 0.6-0.7 | 7-12 |
| <i>Clostridium botulinum</i> , type B, spore | Phosphate buffer, pH 7.0 | 110 | 0.4-1.1 | 7-8 |
| Other microorganisms | | | | |
| <i>Aspergillus</i> sp., conidia | Buffer, pH 4.5 | 55 | 2 | 3.5-4 |
| <i>Aspergillus</i> sp., ascospores | Buffer, pH 4.5 | 75 | 2 | 6-8 |
| <i>Saccharomyces cerevisiae</i> , vegetative | Buffer | 60 | 1 | 5.0 |